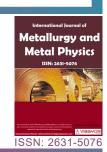


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A Dual Sono/Photochemical Fabrication of the Sage Leaves Extract Coated Silver Nanoparticles



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Abstract

In this study, we prepared the biodegradable sage leaves extract coated silver nanoparticles (SLE-AgNPs) using a green dual sono/photochemical synthesis route. The SLE-AgNPs were successfully synthesized from silver nitrate using the SLE as a green reducing and stabilizing agent. The morphological characterization and functional groups of SLE and SLE-AgNPs were determined by different techniques such as scanning electron microscopy (SEM) with energy-dispersive X-ray (EDX) analyzer, transmission electron microscope (TEM), Fourier transform infrared (FTIR) spectroscopy, Brunauer-Emmett-Teller (BET), dynamic light scattering (DLS), and X-ray diffraction (XRD). We examined the effects of experimental parameters such as the amplitude, sonication time, and temperature on the rheological results. According to the obtained experimental results, we proved that the SLE NPs had a uniform spherical shape under the optimum experimental conditions. SLE-AgNPs were obtained with high colloidal stability. The average diameter range of SLE-AgNPs was ~120 nm while the surface area determined by the BET measurement was 150.67 m2/g. Furthermore, we performed to evaluate the accuracy of the experimental results obtained error analysis using surface tension and conducted the error analysis to assess the accuracy of the experimental results using the surface tension and process parameters. Consequently, we proved that the SLE-AgNPs can be successfully synthesized using the ultrasonic method and the synthesized AgNPs can be a promising nanocarrier in multiple applications for the future.

Keywords

Sage leaves, Silver nanoparticles, Dual sono/photochemical synthesis, Ultrasonic method

Introduction

Nanomaterials have been attracted scientists in various application areas such as textile, sensor,

energy, antibacterial, and drug delivery system due to their superior chemical, biological, optical and mechanical properties [1-3]. It ensures that nano-

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materials have superior properties compared to conventional materials, depending on their size, shape, particle distribution, chemical composition, stability, surface area, and surface properties with more reactivity towards chemicals [4,5]. In recent years, many studies have been reported for the preparation and biomedical applications of metal based nanoparticles such as gold, silver, titanium, and copper [6-10]. Among these nanostructures, silver nanoparticles that have antibacterial properties have been widely studied for their superior physicochemical, biological, optical, and mechanical properties [11-15].

Silver nanoparticles have been reported to be synthesized by different methods such as microwave-assisted method [16], laser ablation method [17], hydrothermal synthesis [18] high energy ball milling [19], chemical reduction method [20], electrochemical synthesis [21], biomimetic synthesis [22], ultraviolet-visible light irradiation [23], and ultrasonic-assisted method [24]. While fabricating silver nanoparticles, most of the methods have some disadvantages, such as the use of toxic volatile organic solvents, high temperature and pressure requirements, toxic by-products, and high-cost [25].

In this study, considering the current disadvantages, we aimed to synthesize the sage leaves extract coated with silver nanoparticles (SLE-AgNPs) by the ultrasonic method in a short time (5 min), economically, without the need for high temperature and pressure with the green chemistry approach [26]. Considering these environmental concerns, the aim of this study is to develop the preparation of SLE-AgNPs with homogeneous distribution by a fast method under high-intensity ultrasonic irradiation based on green chemistry principles. There have been several studies focused on the different bio-materials based matrix such as acalypha fruticosa leaf extracts [27], pergularia daemia leaf extract [28], plant leaf [29], brown algae [30], and green tea [31] extracts acted as a reducing and stabilizing agent to obtain the AgNPs. The functional groups such as ketone, aldehyde, amide, terpenoid, flavone and carboxylic acid in the biomaterial extracts, which have excellent biocompatibility and biodegradability properties and play a dual role in the reduction and stabilization of Ag⁺ to Ag⁰ [32-34]. In the literature review, it was determined that the synthesis and application areas of silver nanoparticles based on plant extract were scattered over a wide spectrum, but there were limited studies in which silver nanoparticles were used in a shorter time and less amount. Lee, et al. investigated the catalytic activity of graphene oxide grafted gold nanoparticles and silver/silver chloride nanoparticles which prepared using green-synthesized by a Portulaca oleracea extract [35]. Huong, et al. observed the antibacterial activity of silver nanoparticles using Sapindus mukorossi fruit pericarp extract [36]. Karadirek, et al. preferred the ultrasound assisted green synthesis for silver nanoparticle attached activated carbon for levofloxacin adsorption [37]. Ahn, et al. reported the development of anticancer properties of silver nanoparticles which were prepared by plant extracts [38]. Albeladi, et al. prepared the silver nanoparticles using salvia officinalis leaf extract using the solution method to show its antimicrobial activity against staphylococcus aureus [25]. In this study, our scientific goal is to determine the effect of the surface tension and process parameters on the rheology characteristic of the synthesized SLE NPs. The attractive effect of our study is that, due to sonication-based cavitation, we have shown that the synthesized nanostructures, have a homogeneous distribution, can be obtained by using a low amount of silver solution for the reaction time of 5 min.

Materials and Methods

Materials

The sage leaves (Salvia officinalis L.) were collected from Antalya (Turkey). Voucher specimen was stored in the Herbarium at the Faculty of Pharmacy, Hacettepe University, Turkey (HUEF 08014). The chemical composition of Sage leaves (Salvia officinalis L.) consisted of the polyphenolic compounds (phenolic acids, polyphenols, flavonoids, phenolic terpenes) and antioxidant constituents (carnosol, rosmadial, carnosine acid, rosmarinic acid and epirosmanol). Sodium hydroxide (NaOH) was purchased from Merck Company. Silver nitrate $(AgNO_3)$ (ACS reagent, \geq 99.0%, molecular weight: 169.87 g mol⁻¹) purchased from Sigma Aldrich Company. We used the sterile syringe filters (Whatman 6900-2502 GD/X 25) which had a 25 mm filter, 0.22-micron retention, and polyvinylidene difluoride (PVDF) membrane to prepare the nanostructure. Samples were stored in sterile polypropylene containers until use. The distilled water had a resistivity of 18.24 M Ω cm at 25 °C. All chemicals and reagents were used without further purification.

Preparation of the sage leaves extract

The fresh sage leaves were collected from Antalya (Turkey) in June and washed with distilled water for several times. They were dried in the vacuum oven until it reached a stable weight at 50 °C. 10g of the dried sage leaves were added into 450 ml of distilled water for 7 days at room temperature in the dark medium without stirring. It was filtered using sterile syringe filters to obtain the SLE at room temperature and the yellow-colored extract was obtained. All samples were stored in sterile polypropylene containers until use at 25 °C.

The dual sono/photochemical fabrication of the SLE-AgNPs

The dual sono/photochemical synthesis route of SLE-AgNPs consisted of four steps. In the first step, 0.84g of AgNO₃ was dissolved in 500 ml of distilled water and stirred for 10 min at room temperature by a magnetic stirrer. 0.4g of NaOH was dissolved in 250 ml of distilled water and stirred for 10 min at room temperature by a magnetic stirrer. In the second step, 100 ml of the sage leaves extract was mixed with the 20 ml of AgNO₃ solution and sonicated for 5 min at 30% amplitude - frequency. In the third step, 2 ml of NaOH solution was added slowly to this solution. By changing the color yellow to brown, the synthesis of SLE-AgNPs based was confirmed (pH = 6) at 25 °C. Finally, SLE-AgNPs were filtered using sterile syringe filters and stored in sterile polypropylene containers until use at 25 °C.

Characterization

We used different advanced techniques such as scanning electron microscopy (SEM) - energy-dispersive X-ray (EDX), transmission electron microscope (TEM), Fourier transform infrared (FTIR) Brunauer-Emmett-Teller spectroscopy, (BET), dynamic light scattering (DLS), and X-ray diffraction (XRD) to characterize the nanostructure. The SEM-EDX (FEI QUANTA, SEM 450) (TedPella, double-coated, 8 mmW × 20 mL) was used to determine the morphology of the structure (6-10 mm working distance, 0-130 Pa pressure, and voltage of 7-10 kV) under low vacuum medium. FTIR (Perkin Elmer, Spectrum Two) (KBr powder and in the 4000-400 cm⁻¹ frequency range with a resolution of 4 cm⁻¹ and 8 scans) was used to observe the functional groups of the samples. Transmission electron microscope (TEM) (Hitachi HighTech HT7700) was used to visualize the nanostructure. BET (ASAP2020 model) was used to determine the specific surface area of the nanostructure. DLS measurements were performed using a Malvern Zetasizer Nano ZS° instrument with 12 runs and were recorded at 25 °C. The surface tensions were performed using a force tensiometer (K11, KRUSS) under different experimental factors such as the amplitude, sonication time, and temperature.

Designation of factors for the multiple-responsive optimization

There are limited reports on the surface tensions of nanostructures in the literature. In this study, we focused on the design of the nanostructure for the multiple-responsive optimization using the rheological measurements of the SLE-AgNPs. These measurements based on the relationship between surface tensions of the samples with different experimental factors such as the amplitude (20-40%), sonication time (1-5 min) and temperature (25-45 °C) were performed in three replicates.

Error analysis

In this study, we calculated different error analysis methods for SLE-AgNPs to evaluate the accuracy of the experimental results obtained the error analysis using surface tension results. These methods were the sum of squares of errors (SSE), the hybrid fractional error function (HYBRID), and the average relative error (ARE) all expressions and given in Table 1 (Equation 2.1-2.3) [39-41].

Stability of the SLE-AgNPs over time

The storage stability of Ag-PVP NPs was evaluated at times 0, 8, 12, and 24 weeks using the UV-vis spectrophotometer and force tensiometer. Four sets of sterile centrifugal tubes containing SLE-Ag-NPs were prepared for each nominal size at 25 °C and all samples were analyzed. The experimental measurements were performed in triplicate.

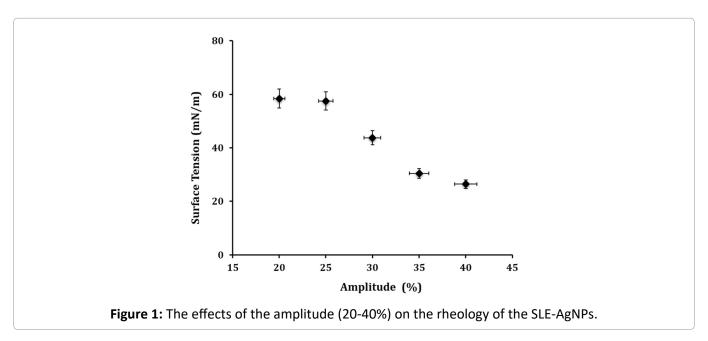
Results and Discussion

In this study, we investigated the morphology and functional groups of the synthesized SLE-Ag-NPs using different techniques such as SEM-EDX, TEM, FTIR, BET, DLS, and XRD. Also, we evaluated the effects of experimental factors such as the amplitude (20-40%), sonication time (1-5 min), and temperature (25-45 °C) on the rheological results. The experimental results presented the optimization of the novel nanostructure for the rheological

Expressions	Error analysis methods	Equation	Ref
$\sum_{i=1}^{n} (\gamma_{cal} - \gamma_{e})^{2}_{i}$	SSE	(2.1)	[39]
$\frac{100}{n-p} \sum_{i=1}^{n} \left[\frac{(\gamma_e - \gamma_{cal})^2_i}{\gamma_e} \right]$	HYBRID	(2.2)	[40]
$\frac{100}{n-p} \sum_{i=1}^{n} \left \frac{\gamma_e - \gamma_{cal}}{\gamma_e} \right _{i}$	ARE	(2.3)	[41]

Table 1: The formulas of the error methods.

Where, n: Number of data points, p: Number of parameters, γ_e : Experimental surface tension and γ_{cal} : Calculated surface tension.



parameters with a focus on surface tensions.

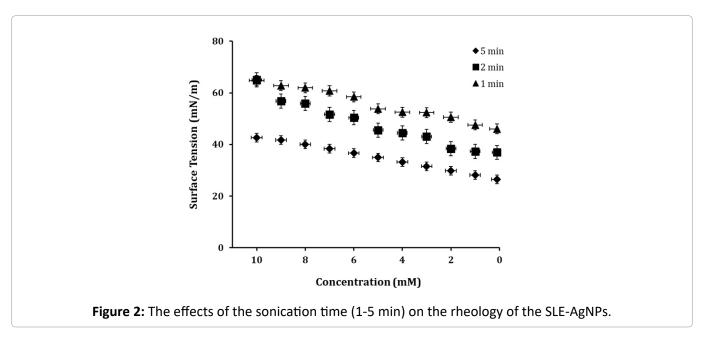
Rheological characterization of the SLE-AgNPs

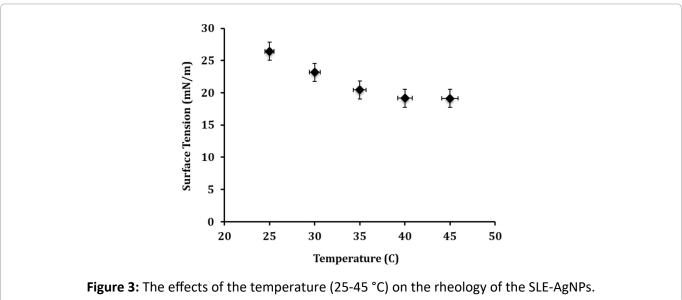
Rheological measurements have a major role in quality control during manufacturing, processing, and characterization of the structures [42,43]. As known, surface tension is an important criterion in the preparation of homogeneous formulations. We focused on the influences of experimental factors on the complex design which plays an important role in the fabrication of AgNPs with rheology measurements. We highlighted the applications of amplitude, sonication time and temperature as a rheological modifier on the surface tension. Also, we reported the optimization of ultrasonic conditions for the fabrication of SLE NPs. For this purpose, we compared experimentally the effects of the amplitude (20-40%), sonication time (1-5 min),

and temperature (25-45 °C) on the rheology of the SLE-AgNPs and these results were given in Figure 1, Figure 2 and Figure 3, respectively.

The surface tensions of SLE/solvent from 58.45 mN/m to 26.43 mN/m with changing the amplitude of the process from 20-40% at 25 °C were recorded for SLE-AgNPs (Figure 1). According to the results, we observed that the surface tension of SLE/solvent increased with the increase of the value of the amplitude. We proved that the surface tension changed at different amplitude values due to the intensity and frequency of the sonication. (Figure 1) [44].

The surface tensions of SLE/solvent from 42.67 mN/m to 26.43 mN/m with changing concentration of the SLE-AgNPs from 10-0.1 mM at 25 °C were recorded for 5 min. The surface tensions of SLE/sol-





vent from 64.99 mN/m to 36.91 mN/m with changing concentration of the SLE-AgNPs from 10-0.1 mM at 25 °C were recorded for 2 min. The surface tensions of SLE/solvent from 64.77 mN/m to 46.04 mN/m with changing concentration of the SLE-AgNPs from 10-0.1 mM at 25 °C were recorded for 1 min. According to the experimental results, we observed that the surface tensions of SLE/solvent increased with the increase of the concentration of SLE-AgNPs. It was proven that the surface tension changed at different sonication times depending on the AgNPs increased (Figure 2) [45,46].

In Figure 3, the surface tensions of SLE/solvent from 26.43 mN/m to 19.12 mN/m with changing the temperature (25-45 °C) of the SLE-AgNPs were recorded. We observed that the surface tensions

of SLE/solvent increased with the increase of the concentration of SLE-AgNPs. The effects of the amplitude, sonication time, and temperature on the surface tension of the nanostructure produced using the sonication method owing to the collapse of microbubbles in the nanostructure irradiated with ultrasound was experimentally examined in this study [47,48]. Accordingly, the sonochemical effects on the surface tension of the SLE mediated AgNPs owing to the ultrasound cavitation in the nanostructure were proven. To our knowledge, this study is the first to investigate the effects of the sonochemical method on the rheology with the SLE mediated AgNPs.

Characterization of the SLE-AgNPs

In this study, we presented the development of

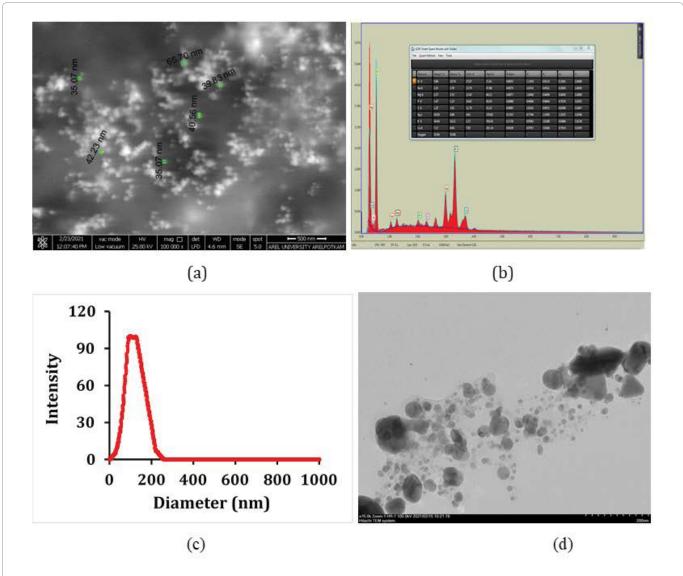


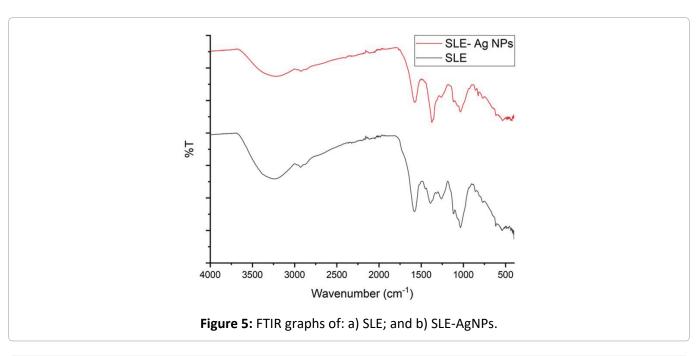
Figure 4: a) The SEM image; b) The EDX image; c) The TEM image; and d) DLS results of the SLE-AgNPs.

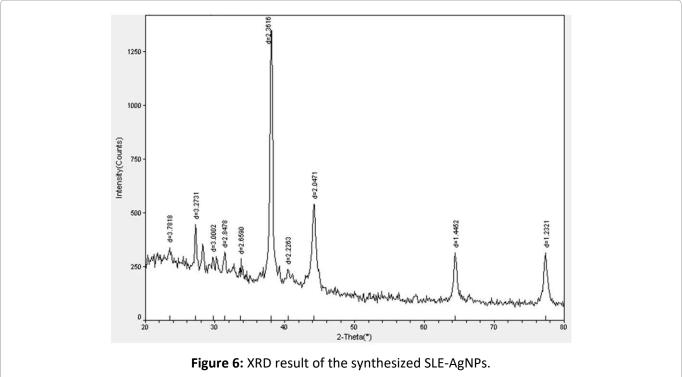
the dual sono/photochemical synthesis route using the SLE matrix as a phytochemical under the ultrasonic irradiations Ag⁺ into AgNPs.

The surface morphology of synthesized SLE-Ag-NPs was performed by SEM and EDX techniques. (Figure 4a and Figure 4b). The SEM analysis revealed that the SLE-AgNPs had spherical-shaped particles with a homogeneous distribution. The size of the nanoparticles varied between 35.07 and 65.70 nm. Similar SEM results were obtained for the AgNPs prepared by Chowdhury, et al. [49]. The EDX analysis was performed to determine the elemental composition of the SLE-AgNPs. Figure 4b illustrated the EDX spectrum of the SLE-AgNPs. The results indicated the presence of an intense peak due to the existence of Ag (20.20%), Ca (7.17%), O (33.94%), Na (3.35%), Mg (2.37%), P (1.47%), S (1.25%), and K (24.24%) elements in the biosynthe-

sized SLE-AgNPs. The results of the DLS technique demonstrated that there was a uniform size distribution of the AgNPs with the hydrodynamic average size of 120 nm (PdI: 0.007 and intercept: 0.944) and good dispersion of AgNPs in 10 mM phosphate buffered saline (PBS) (1x pH 7.4) (Figure 4b). TEM image showed that biosynthesized AgNPs had a particle size distribution between 10 and 50 nm (Figure 4c). Thus, the results of the TEM analysis of the AgNPs we obtained were compatible with our SEM results. By using the results of BET measurements, we found that the surface area was 150.67 m²/g.

The FTIR technique of SLE and SLE-AgNPs was used to investigate functional groups in the chemical structure and the interaction between AgNPs and SLE matrix in the region between 400 and 4000 cm⁻¹. FTIR analysis of (a) SLE and (b) SLE-AgNPs





were given in Figure 5. According to the FTIR results, the characteristic peaks of SLE were observed at 3270.68 cm⁻¹ (-OH stretching vibration), 2927.41 cm⁻¹ (C-H stretching vibration), 1585.19 cm⁻¹ (C-O stretching vibrations), and 1031.73 cm⁻¹ (C-O-C vibrations). Furthermore, the characteristic peaks of SLE-AgNPs were observed at 3303.46 cm⁻¹ (-OH stretching vibration), 1577.49 cm⁻¹ (C=O, carbonyl groups), 1373.07 cm⁻¹ (C-H wagging vibrations) and 1035.59 cm⁻¹ (amine groups N-H). According to these results, it was observed that the peaks of

-OH stretching vibration, CO stretching vibrations, and C-O-C vibrations were relatively smaller in the presence of AgNPs, but the relative intensity of the C-H wagging vibrations peak increased. We also observed small peaks at 528 cm⁻¹ due to the polyphenols which showed there are phenolic and carbonyl groups on the surface of the AgNPs. Similar spectra results indicate there was not much change in the SLE and SLE-AgNPs [29,36].

X-ray Diffraction (XRD) result of SLE-AgNPs was performed to investigate the effect of synthesized

Sample Method **Characterization results** Ref. Plant extract - AgNPs Biosynthesis Spherical shape/17 nm 52 [53] Graphene oxide-AgNPs Hybrids Sonochemical synthesis Irregularly shaped/15-35 nm Perilla frutescens Leaf Extract- AgNPs Phytosynthesis Spherical shape/25.71 nm 54 AgNPs in Y-zeolite substrate [55] Sonochemical synthesis Spherical shape/10 nm Colloidal AgNPs [56] Sonochemical synthesis Spherical shape/3.5 nm Purple heart plant leaves extract-AgNPs Biosynthesis Spherical shape/98 nm [57] Myristica fragrans seed extract AgNPs Biosynthesis Spherical shape/5-20 nm [58] Azadirachta indica leaf extract AgNPs Biosynthesis Spherical shape/4-19 nm [59] Biosynthesis Aegle marmelos fruit extract AgNPs [60] [61] Allium ampeloprasum extract AgNPs Biosynthesis Spherical shape/116 nm **SLE-AgNPs** Dual sono/photochemical Spherical shape/17.56 nm This study

Table 2: The comparison of characterization results of the various AgNPs.

AgNPs on the crystalline structure. The results were displayed in Figure 6. Figure 6 showed that characteristic peaks were formed around $2\theta = 38.1^{\circ}$, 44.4°, 64.4°, and 77.4° which corresponding to (111), (200), (220), and (311) planes of AgNPs, respectively (JCPDS file No.00-004-0783) [50,51]. We calculated the average crystallite size of SLE-AgNPs using Equation 3.1 (Debye-Scherer's equation) [51].

$$D = 0.9.\lambda / \beta.\cos\theta \tag{3.1}$$

Where λ : 0.1541 nm, β : The full width at half maximum (FWHM), θ : The Bragg's angle in degrees, and D: The average particle size.

The average crystallite size of AgNPs was measured from the plane (1 1 1) using Eq. 3.1. It was calculated to be around 17.56 nm of SLE-AgNPs. According to our experimental results, the estimated fabrication mechanism of the AgNPs was described in Eqs. 3.2-3.9.

$$H_2O \to {}^*OH + {}^*H$$
 (3.2)

$$^*OH + ^*OH \rightarrow H_3O_3$$
 (3.3)

$$^*H + O_{\gamma} \rightarrow HO_{\gamma}^*$$
 (3.4)

$$HO^{-} + H_{2}O_{2} \leftrightarrow HOO^{-} + H_{2}O$$
 (3.5)

$$HO_2^* + ^*OH \rightarrow H_2O + O_2$$
 (3.6)

$$AgNO_3 \rightarrow Ag^+ + NO_3^- \tag{3.7}$$

$$SLE-Ag^+ + H_2O_2 \leftrightarrow SLE-Ag^0 + H^+ + HOO^*$$
 (3.8)

$$HOO^{-} + (SLE -) Ag^{+} \leftrightarrow HOO^{*} + SLE -Ag^{\circ}$$
 (3.9)

In previous studies, it was proven that bio-based products such as nanostructures obtained from biological systems and products mediated by the vegetable, animal oils, and plant extracts are bio-

degradable. For this purpose, our scientific goal was to prepare the biodegradable plant extract mediated silver nanoparticles without using any toxic reducing agents in this study. The results of the synthesized AgNPs were appreciable from the comparison with the results of previous studies in the literature (Table 2) [52-61].

Error analysis of SLE-AgNPs

In this study, the error analysis results of SLE-Ag-NPs were compared with different error methods such as SSE, HYBRID, and ARE using the surface tension measurements. When Eq. 2.1-2.3 were applied with the repetition of the data obtained from the surface tension measurements, the minimum error results were observed. According to results, we calculated the parameters of SSE, HYBRID, and ARE were 0.12, 0.15, and 0.48, respectively. Accordingly, it was proven with the minimum error results. Although such error calculations were reported in the literature, no study gives the relation of the surface tension for the AgNPs. Also, there are no systematic reports on the comparison of the error methods of the AgNPs in the literature to determine the change in the surface tension of the nanostructure under the sonochemical cavitation.

Stability of SLE-AgNPs

The storage stability of the SLE-AgNPS was evaluated using surface tension measurements and UV-Vis spectroscopy. The storage stability of the SLE-AgNPS was monitored by checking its surface tension and absorbance at 25 °C. Also, the storage stability of the SLE-AgNPs was confirmed by the UV-Vis absorption spectra measured as a func-

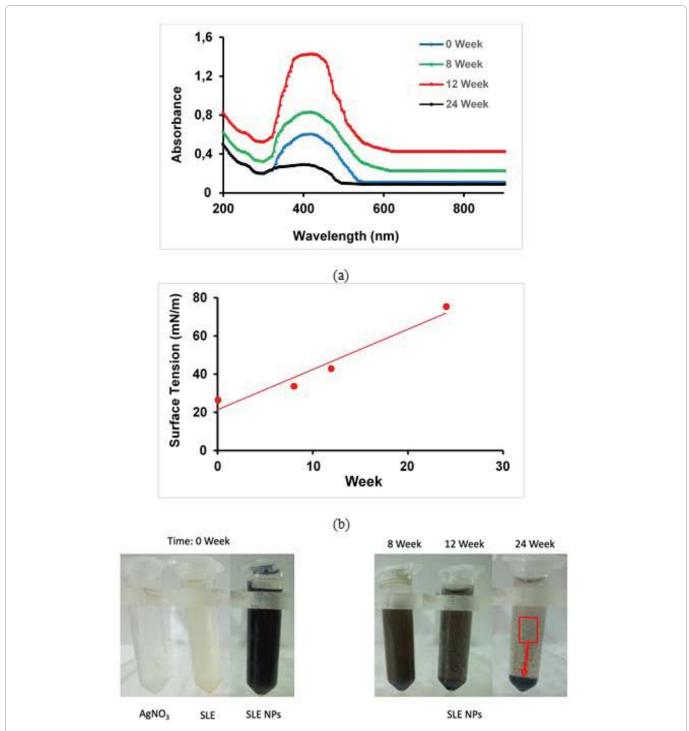


Figure 7: a) The UV-Vis spectra; b) Surface tension of the SLE-AgNPs; and c) The photographs of the silver nitrate solution, SLE, and SLE-AgNPs at different times.

tion of time (0-24 Week). Besides, SLE-AgNPs were proved to have two characteristic peaks at absorption spectra 265 and 465 nm (Figure 7). Furthermore, the surface tension of the SLE-AgNPS after 24 weeks of storage was found to be 75.43 ± 1.0 mN/m (Figure 7). These results exhibited an indication of its excellent stability with its photographs at different times for the SLE NPs [62].

Conclusion

In summary, the SLE-AgNPs were prepared successfully by the green dual sono/photochemical synthesis method and the synthesized SLE-AgNPs displayed sensitivity to factors such as the amplitude (20-40%), sonication time (1-5 min), and temperature (25-45 °C) on the surface tension. The effective experimental factors were calculated to

determine the optimum condition of the preparation of the uniform nanostructure with compatible error results. The characterization results showed that the SLE-AgNPs had a uniform spherical-shaped structure with a large surface area (150.67 m²/g) and have potential in various applications of scientific and industrial fields in the future.

Declaration of Competing Interest

The authors declare no conflict of interests.

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